A study of inactivation of *Chlamydia trachomatis* by normal human serum

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SUMMARY To assess the effect of human serum on the viability of *Chlamydia trachomatis*, organisms were mixed with unheated and heat inactivated homologous serum, and the numbers surviving after incubation at 37°C for 1 hour were compared. With a pool of sera obtained from 12 donors, the number of chlamydiae surviving incubation in unheated serum was less than 1% of that surviving incubation in heat inactivated serum. The antichlamydial activity of the unheated pooled serum samples could be noticeably reduced by treatment with Mg-EGTA (ethyleneglycol-bis (β-amino ethyl ether)-N,N'-tetra-acetic acid). This indicated a requirement for calcium ions and showed that the alternative pathway of complement activation played only a minor role, if any, in the inactivation process. When 12 serum samples were tested individually it was found that four inactivated chlamydiae to an extent comparable with that seen with the pooled serum. The other eight samples showed only moderate (or slight) antichlamydial activity, with survival rates in unheated serum of 20-60% (or more than 60%) of those in heat inactivated serum. There was no correlation between the titres of antichlamydial antibodies and antichlamydial activity, all serum samples having undetectable or low concentrations of antibody on measurement by micro-immunofluorescence. The antichlamydial activity destroyed by heating was restored, however, when heat inactivated serum was mixed with an equal volume of an unheated serum that was not inhibitory to chlamydiae. When the latter serum was heated before addition antichlamydial activity was not restored, indicating the requirement of both a heat stable and a heat labile factor. This observation and the need for calcium ions for inactivation of chlamydiae are compatible with killing mediated by antibody and complement. Thus serum samples from individuals with no clinical or serological evidence of infection with chlamydiae vary in their ability to inactivate the organism, some having antichlamydial activity which is possibly mediated by antibody and complement.

Introduction

Although *Chlamydia trachomatis* is an important human pathogen,¹ little is known of the factors that determine its pathogenicity. The virulence of many micro-organisms is determined by their surface structure, with cell wall or capsular antigens acting to promote colonisation or to resist host defence mechanisms.² The cell wall of *C trachomatis* is

structurally and chemically similar to that of Gram negative bacteria, with an inner cytoplasmic membrane surrounded by a trilaminar outer membrane.³ All *Chlamydia* spp also contain a group antigen consisting of lipopolysaccharide, the immunodominant determinant of which is similar to, but not identical with, 2-keto-3-deoxyoctanoic acid from salmonella lipopolysaccharide.⁴ Some strains of Gram negative bacteria are susceptible to the bactericidal activity of normal human serum which may be mediated by antibody and complement, although bacterial lipopolysaccharide may activate the alternative complement pathway in the absence of antibody.⁵ In view of the similarity between the

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cell walls of *C trachomatis* and of Gram negative bacteria, we investigated the effect of normal human serum on the viability of chlamydiae with particular regard to the possible role of the classic and alternative complement pathways, as activation of complement by either mechanism may contribute to immunity from chlamydial infection and development of disease.

Materials and methods

CHLAMYDIA TRACHOMATIS

A strain of *C trachomatis* designated 6234 was used. It had been isolated from the urethra of a patient with non-gonococcal urethritis and was passed four times in McCoy cells treated with cycloheximide⁶ before being stored in liquid nitrogen in sucrose phosphate medium (2SP) containing 10% heat inactivated fetal calf serum.

SERUM SAMPLES

Blood samples were obtained by venepuncture from volunteers on the staff of the Clinical Research Centre after approval had been given by the Ethics Committee. This group of subjects was chosen in the expectation that most of the serum samples would have low or undetectable titres of chlamydial antibodies. Each blood sample was allowed to clot for about 1 hour at room temperature and the serum was separated by centrifugation and stored in small volumes at -70°C . Complement assays (kindly performed by Mrs R Hill, Division of Immunological Medicine, CRC) showed that all the serum samples had normal levels of haemolytic complement activity.

In preliminary experiments a pool containing serum from 12 donors was used. Such pooled samples have been used previously to study the sensitivity to serum of another genital pathogen, *Neisseria gonorrhoeae*.⁷ In later experiments serum samples from 12 individual donors, six of whom had contributed to the serum pool, were used separately.

Mg-EGTA

A stock solution of 100 mmol/l Mg-EGTA (ethylene-glycol-bis (β -amino ethyl ether)-N,N'-tetra-acetic acid) was prepared in physiological saline as described by Fine *et al.*⁸ Mg-EGTA selectively inhibits the classical pathway of complement activation by chelating calcium ions, but does not inhibit the alternative complement pathway.

TEST SYSTEM

To determine whether *C trachomatis* organisms were inactivated by serum, 20 μl of a chlamydial suspension were added to 150 μl of serum supplemented with 30 μl of either physiological saline

or phosphate buffered saline (PBS). In some experiments the 30 μl of saline were replaced by 20 μl of the stock solution of Mg-EGTA with 10 μl of saline.

After addition of chlamydiae to the serum samples, the mixtures were incubated at 37°C for one hour. The number of viable chlamydiae in the mixtures at the end of the incubation period was determined by titration in McCoy cells treated with cycloheximide using centrifuge assisted inoculation.⁶ The cell monolayers were coded before microscopic examination to avoid subjective bias. Inactivation of chlamydiae was assessed at the end of the incubation period by comparing the number of viable chlamydiae in unheated serum with the number in homologous serum that had previously been heated at 56°C for 30 minutes to inactivate complement.

SEROLOGICAL TEST

The presence of IgM and IgG antibodies to *C trachomatis* in the serum samples was determined by microimmunofluorescence.⁹

Results

INACTIVATION OF *C TRACHOMATIS* BY POOLED HUMAN SERUM AND EFFECT OF ADDING Mg-EGTA

In six experiments, the results of one of which are shown in table I, the number of viable chlamydiae

TABLE I *Survival of C trachomatis after incubation at 37°C for 1 hour in unheated and heat inactivated pooled human serum with and without addition of Mg-EGTA*

Treatment of serum	No of chlamydiae recovered*
Unheated	2.21
Heat inactivated	4.45
Unheated with 10 mmol/l Mg-EGTA	4.32
Heat inactivated with 10 mmol/l Mg-EGTA	4.61

*Inclusion forming units/ml ($\log_{10}$).

recovered after incubation in unheated serum was less than 1% of the number recovered after incubation in heat inactivated serum. The serum was supplemented with 10 mmol/l Mg-EGTA in four experiments, in each of which the inactivation of chlamydiae by unheated serum was markedly inhibited. In two experiments the inactivation of chlamydiae was completely inhibited, while in the other two survival of the organisms in the Mg-EGTA treated unheated serum was about 70% of that in heat inactivated serum as opposed to less than 1% survival in the untreated serum (table I). This

indicated that the inactivation of chlamydiae by serum predominantly involved the classical complement pathway, with only a possible minor contribution by the alternative complement pathway.

ANTICHLAMYDIAL ACTIVITY AND TITRES OF CHLAMYDIAL ANTIBODY IN INDIVIDUAL SERUM SAMPLES

In view of the inactivation of chlamydiae by pooled serum, we investigated the effect of individual serum samples on the viability of *C. trachomatis*. Of 12 serum samples, four inactivated chlamydiae to an extent comparable with that seen with pooled serum, whereas eight did so only slightly (figure). Survival of

chlamydiae in three of these eight serum samples was generally 20-60% of that in heat inactivated homologous samples, while the other five generally allowed more than 60% of the survival rate in heat inactivated serum. There was, however, no correlation between the antichlamydial activity of the individual serum samples and the antibody titre measured by microimmunofluorescence (table II).

TABLE II *Antichlamydial activity and antibody titres of individual human serum samples*

Serum No	Antichlamydial activity*	Titre† of anti-chlamydial antibody	
		IgM	IgG
1	slight (>60%)	—	1/1 (F)
2	slight (>60%)	—	1/1 (F)
3	slight (>60%)	1/1 (F)	1/2
4	slight (>60%)	—	—
5	slight (>60%)	—	1/8 (F)
6	moderate (20-60%)	—	—
7	moderate (20-60%)	—	1/1 (F)
8	moderate (20-60%)	—	—
9	great (<2%)	—	—
10	great (<2%)	—	1/2 (F)
11	great (<2%)	—	1/2
12	great (<2%)	1/2	1/2

*Survival of chlamydiae in unheated serum expressed as a percentage of survival in heat inactivated homologous serum.

†Highest dilution of serum showing fluorescence.

(F) = faint fluorescence.

— = no antibody detected.

REQUIREMENT OF A HEAT LABILE SERUM FACTOR FOR INACTIVATION OF CHLAMYDIAE

As described above, the ability of pooled human serum or of individual serum samples to inactivate chlamydiae was abolished by heating, thus suggesting a possible requirement for complement. This idea was supported by the observation that the antichlamydial activity of heat inactivated serum could be restored by adding an equal volume of an unheated serum that was not itself inhibitory to chlamydiae (table III). Furthermore, heating of the

TABLE III *Survival of C. trachomatis after incubation for 1 hour at 37°C in human serum*

Serum No*	Treatment of serum	No† of chlamydiae recovered
9	unheated	<1.00
9	heat inactivated	4.03
1	unheated	4.06
1	heat inactivated	3.99
9 plus 1	heat inactivated } unheated	2.20
9 plus 1	heat inactivated } heat inactivated	4.03

*Refers to serum samples listed in table 2.

†Inclusion forming units/ml (log₁₀).

FIGURE *Survival of chlamydiae in heat inactivated and unheated human serum samples after incubation at 37°C for 1 hour. Homologous heat inactivated and unheated serum samples are joined by lines.*

donor serum prior to the addition failed to restore the antichlamydial activity of the recipient heat inactivated serum. This showed that the inactivation of chlamydiae by serum required both a heat stable and a heat labile factor.

Discussion

The results presented here show that *C trachomatis* is susceptible to inactivation by unheated human serum. The observation that this was markedly inhibited when calcium ions were chelated showed that the alternative complement pathway played at best only a minor role in the inactivation process. Inactivation mediated by serum required heat stable and heat labile factors and calcium ions, which suggests involvement of the classical complement pathway. Although there was no correlation between the antichlamydial activity of serum and the level of detectable antichlamydial antibody, this does not rule out the possibility of killing mediated by antibody. The antibody detected by microimmunofluorescence in serum samples that did not inactivate chlamydiae may have been directed against chlamydial antigens that did not function as efficient triggers of lysis mediated by complement. An example is seen with *N gonorrhoeae* where antibody to the lipopolysaccharide may kill the bacteria in the presence of complement, but antibodies to pili do not.¹⁰ A further consideration is that antibody responsible for complement activation may have been present at a concentration too low to be detected by microimmunofluorescence but was nevertheless capable of killing chlamydiae because of the amplification effect of the complement cascade. Serum samples that appear to lack antichlamydial antibody when assayed by microimmunofluorescence may contain antibody detectable by a more sensitive immunoelectrophoretic transfer method.¹¹ A full understanding of the mechanism of serum inactivation of chlamydiae needs more information on

the immunochemical and structural composition of the chlamydial cell surface, together with the application of sensitive assays for detecting antibodies directed against individual antigens.

In many serological studies chlamydial antibody titres of $<1/8$, measured by microimmunofluorescence, are regarded as normal. The results of this study, however, indicate that serum with such titres may inactivate chlamydiae. Whether this is important in terms of development of disease and immunity to infection remains to be determined.

References

1. Taylor-Robinson D, Thomas BJ. The role of *Chlamydia trachomatis* in genital-tract and associated diseases. *J Clin Microbiol* 1980; **33**:205-33.
2. Smith H. Microbial surfaces in relation to pathogenicity. *Bacteriological Reviews* 1977; **41**:475-500.
3. Schachter J, Caldwell HD. Chlamydiae. *Annu Rev Microbiol* 1980; **34**:285-309.
4. Kuo C-C, Caldwell HD, Wang S-P, Grayston JT. Antigens of *Chlamydia trachomatis*. In: Hobson D, Holmes KK, eds. *Non-gonococcal urethritis and related infections*. Washington DC: American Society for Microbiology, 1977; 176-85.
5. Schreiber RD, Morrison DC, Podack ER, Müller-Eberhard HJ. Bactericidal activity of the alternative complement pathway generated from 11 isolated plasma proteins. *J Exp Med* 1979; **149**:870-82.
6. Thomas BJ, Evans RT, Hutchinson GR, Taylor-Robinson D. Early detection of chlamydial inclusions combining the use of cycloheximide-treated McCoy cells and immunofluorescence staining. *J Clin Microbiol* 1977; **6**:285-92.
7. Johnson AP, Osborn MF, Taylor-Robinson D. Iron-mediated protection of *Neisseria gonorrhoeae* against killing by normal human serum. *FEMS Microbiol Letters* 1978; **4**:225-7.
8. Fine DP, Marney SR, Colley DG, Sergeant JS, Des Prez RM. C3 shunt activation in human serum chelated with EGTA. *J Immunol* 1972; **109**:807-9.
9. Thomas BJ, Reeve P, Oriel JD. Simplified serological test for antibodies to *Chlamydia trachomatis*. *J Clin Microbiol* 1976; **4**:6-10.
10. Ward ME, Lambden PR, Heckels JE, Watt PJ. The surface properties of *Neisseria gonorrhoeae*: determinants of susceptibility to antibody complement killing. *J Gen Microbiol* 1978; **108**:205-12.
11. Newhall WJ, Batteiger B, Jones RB. Analysis of the human serological response to proteins of *Chlamydia trachomatis*. *Infect Immun* 1982; **38**:1181-9.