Detection of human papillomavirus types in balanitis xerotica obliterans and other penile conditions

P W Y Lau, N Cook, H Andrews, A Bracka, S H Myint

Abstract

Objectives—To determine the prevalence of human papillomavirus (HPV) types 6, 11, 16 and 18 in foreskin biopsies from patients with balanitis xerotica obliterans (B XO) and other penile conditions.

Materials and methods—Foreskin biopsy specimens from 24 patients with penile lesions and 5 control patients were analysed by type-specific polymerase chain reaction (PCR).

Results—HPV6 or HPV16 were not detected in patients with B XO. HPV6 was detected in 2 controls.

Conclusions—Genital papillomaviruses do not have a strong association with B XO.

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Keywords: Balanitis xerotica obliterans; Papillomavirus

Introduction

Balanitis xerotica obliterans (B XO), also known as lichen sclerosis of the penis is an unusual condition that predominantly affects young adults and children. It is first manifest as white atrophic patches which may be mildly itchy. The plaques form bands which will eventually start to constrict the penis. The patient may suffer recurrent episodes of balanitis and eventually circumcision is often necessary.1 Squamous cell carcinoma has also been reported as a rare complication of the condition.2 The aetiology of this condition is unknown although genetic, autoimmune, hormonal and infectious mechanisms have been proposed.4 The search for a possible infectious agent has led us to examine the presence of known genital viruses in this condition. We present in this paper the results of screening for specific genital papillomaviruses, a possibility raised by the association of these viruses with squamous carcinoma.

Materials and methods

Human papillomaviruses (HPV)-specific PCR Oligonucleotide primers were synthesised from published sequences for HPV types 6, 11, 16 and 18. The sequences were derived from the E6 region and are shown in table 1.

Table 1 Oligonucleotide primer sequences used in the PCR amplification (from reference 5). All primers are annotated with reference to Genbank sequences and are written in a 5' to 3' orientation

<table>
<thead>
<tr>
<th>HPV type</th>
<th>5' Primer</th>
<th>3' Primer</th>
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<tbody>
<tr>
<td>HPV6</td>
<td>TCT ATC TAT GCA TAC GTT GC</td>
<td>CCA TTT TGT ATA TGA TTO GT</td>
</tr>
<tr>
<td>HPV11</td>
<td>TCT TTT TTT GCA CAC TCT GC</td>
<td>GAC GTA TTT TOA TTT ATG GC</td>
</tr>
<tr>
<td>HPV16</td>
<td>CAG GAC CCA CAG GAC GGA CC</td>
<td>CCC AGC AAG CTC GGC GAC TA</td>
</tr>
<tr>
<td>HPV18</td>
<td>GCT TTT AGG ATC CAA CAC GC</td>
<td>GGT CAG GGT AAG CAC GAC GT</td>
</tr>
</tbody>
</table>
Extracted DNA (0.5 μl) was amplified using the appropriate primers in an Amplitaq assay (Perkin-Elmer, Norwalk, USA) using the recommended buffer and reagent concentrations, and 5 units of Amplitaq polymerase. Appropriate positive controls, consisting of HPV insertion sequences derived from plasmids (kindly supplied by Professor E.M de Villiers of the DeutschesKrebsForschung-Zentrum, Heidelberg) were used in all amplification runs. A negative control, sterile water, was also included and precautions were taken to minimise the risks of DNA carryover. After an initial “hot-start” of 94°C for 10 minutes, different cycling parameters were used for specific virus types. For amplification of HPV6 and HPV11 sequences, thirty cycles of 95°C for 30 seconds, 58°C for 30 seconds and 72°C for one minute were followed by a final extension at 72°C for 10 minutes. For amplification of HPV16 and HPV18 sequences the thermal cycling protocol was thirty cycles of 95°C for 30 seconds, 68°C for 30 seconds and 72°C for one minute, with a final extension at 72°C for 10 minutes. All amplifications were undertaken in a Geneamp 9600 cycler (Perkin-Elmer, Norwalk, USA). Amplification products were visualised by electrophoresis in 2% agarose gels and ethidium bromide staining with ultraviolet illumination.

**Results**

Figures 1 and 2 show the results of positive (typeable) specimens. All the specimens were β-globin PCR positive. Table 2 summarises the results and histological diagnoses. It can be seen that none of the BXO specimens had detectable HPV sequences. The genital warts had detectable HPV as might be expected but interestingly only one was a common genital type. One of the cases of non-specific inflammation had detectable HPV but without follow up clinical information we cannot assess any pathological significance. Two of the five control specimens were positive for HPV6 only (data not shown).

**Discussion**

The evidence for an infectious aetiology for BXO is predominantly anecdotal. There is some support for the hypothesis, however, from studies that suggest there is chronic antigen stimulation in the disease. The association with squamous carcinoma might suggest an agent that had oncogenic potential. Much evidence has accumulated that specific HPV...
types have this potential. One of these is HPV16 but this was not found in our patients using a very sensitive technique. The numbers are small but statistically the prevalence of HPV types in the BXO and non-BXO groups is different ($p < 0.05$).

The prevalence of genital human papillomavirus infections in the adult male population is not known with any great precision but studies in a subset, those that are sexually active have suggested that between 20 and 45% of men would be expected to carry genital papillomaviruses. This figure is higher in contacts of women with known cervical HPV infection. We do not know the sexual histories of the patients enrolled in this study but it is clear that the prevalence in our control patients is consistent with other studies but the prevalence in patients with BXO is significantly less. This would negate a significant or consistent role for human genital papillomaviruses in balanitis xerotica obliterans. The search for an infectious agent should continue.

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