Magnetic resonance imaging, thallium-201 SPET scanning, and laboratory analyses for discrimination of cerebral lymphoma and toxoplasmosis in AIDS

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Objectives: To compare the results of magnetic resonance imaging (MRI) and thallium-201 (201Tl) SPET scanning with laboratory analyses including CSF DNA detection, brain biopsy, and necropsy in the discrimination of cerebral lymphoma and toxoplasmosis in patients with AIDS.

Methods: A retrospective study of 32 patients infected with HIV who had focal CNS lesions on MRI as a result of either lymphoma or toxoplasmosis.

Results: 18 patients had lymphoma, 12 had toxoplasmosis, and two had both. Toxoplasma IgG antibodies were detected in only seven patients—four with toxoplasmosis, two with lymphoma, and one with both diagnoses. Epstein–Barr virus DNA was detected in CSF of all six patients with lymphoma and none of two with toxoplasmosis. MRI showed multiple lesions in 23 patients, appearances did not discriminate between lymphoma and toxoplasmosis; nine patients had single lesions, of these eight had lymphoma (p=0.044, two tailed Fisher’s exact test) 201Tl SPET showed accumulation in 17 with lymphoma and six with toxoplasmosis (p = 0.034, two tailed Fisher’s exact test). Of nine patients with single lesions on MRI and 201Tl SPET with focal accumulation eight had lymphoma. 201Tl SPET uptake ratios of ≥2.9 were only seen with lymphoma.

Conclusion: Knowledge of patients’ toxoplasma serostatus does not aid discrimination between lymphoma and toxoplasmosis. Single lesions on MRI with focal accumulation of 201Tl strongly suggest lymphoma. Multiple lesions on MRI with 201Tl SPET uptake ratios ≥2.9 also suggest lymphoma; uptake ratios less than 2.1 do not aid discrimination. Detection of Epstein–Barr virus DNA in CSF is highly sensitive and specific for cerebral lymphoma.

Keywords: lymphoma; toxoplasmosis; magnetic resonance imaging; thallium-201; SPET

Introduction
Neurological dysfunction has been reported as a clinical manifestation of AIDS in 30–70% of patients.1, 2 Whereas both human immunodeficiency virus (HIV) itself and opportunistic infection by cytomegalovirus (CMV) may cause diffuse CNS disease,4–6 encephalitis due to Toxoplasma gondii is the most frequent cause of focal CNS disease, accounting for up to 70% of such cases.7 CNS lymphoma, although very uncommon in the general population, occurs in approximately 5% of patients with AIDS in industrialised countries.8 Other causes of focal lesions include cryptococcosis, tuberculous abscesses, and metastatic tumours. The clinical presentation of patients with focal CNS lesions caused either by T gondii or CNS lymphoma is indistinguishable and the neuroradiological abnormalities demonstrated by computed tomography (CT) or magnetic resonance imaging (MRI) frequently fail to discriminate between the two diagnoses.9, 10 Although a tissue diagnosis may be made by a stereotactic brain biopsy most centres do not routinely perform this procedure. Instead, patients with focal CNS lesions are treated empirically with anti-T gondii drugs, a clinical and imaging response to therapy being diagnostic. Brain biopsy is then reserved for those patients who fail to respond to therapy or who have neuroradiological features at presentation that are atypical. Brain biopsy is invasive, requires the availability of a neurosurgical department, and in HIV infected patients has an associated surgical morbidity (up to 12%) and mortality (up to 2%).11 Sampling errors, especially in those with multiple lesions, mean the procedure is non-diagnostic in between 5–33% of cases.12

In order to aid diagnostic discrimination without the need for brain biopsy, alternative diagnostic tests have been used. Detection of Epstein–Barr virus (EBV) DNA in CSF by the polymerase chain reaction (PCR) has been shown to be a highly sensitive and specific technique for diagnosis of CNS lymphoma in AIDS.13, 14 In contrast, detection of T gondii DNA in CSF by PCR has only moderate sensitivity.15 Because of contraindications to lumbar puncture including mass effect, or midline shift, CSF examination is not always possible, thus limiting the diagnostic usefulness of this molecular technique. Alternative imaging methods have been used including MRI spectroscopy, which also fails to discriminate between T gondii abscesses and lymphoma,16 and single photon emission tomography (SPET) using thallium-201 (201Tl).17–19

In the general population 201Tl SPET imaging has been used to localise brain tumours: tumour
upshowsthat201Tl does not accumulate in non-neoplastic lesions such as infection and radiation necrosis. In AIDS patients, 201Tl SPET imaging has been shown in some studies to aid discrimination between CNS lymphoma and T gondii causing focal CNS lesions.

We undertook a retrospective study of 32 HIV positive patients presenting with focal CNS lesions in whom we compared the radiological diagnosis made by MRI and by 201Tl SPET scanning. During the study period a total of 302 HIV infected patients had MRI.

Of the 32 patients studied 29 were men (all were white and homosexual) and three were women (two were white, one was of African origin, all were heterosexual). Their ages ranged from 27 to 53 (median 36) years. CD4 lymphocyte counts ranged from 0.02 × 10^9/l to 0.02 × 10^9/l (median 0.09) to 0.02 × 10^9/l.

At the time of their neurological presentations all patients had been admitted under the care of an HIV/AIDS specialist (RFM) and were also seen by a neurologist (MJGH); they were investigated according to a unit protocol. Following a clinical assessment, investigations included measurements of IgG antibodies to T gondii using a latex agglutination assay (Elkin, Mast Diagnostics). In those with positive results (titre >1:16) samples were sent to the toxoplasmosis reference laboratory for detection of IgG antibodies by the Sabin–Feldman dye test and measurement of IgM antibodies by a particle agglutination assay (Toxo-ISAGA; Bio Mérieux). Note was also made of patient’s use of co-trimoxazole for prophylaxis of Pneumocystis carinii pneumonia (PCP).

Cranial MRI was performed at 1.5 Tesla on a standard clinical system (Siemens Magnetom 6 SP, Erlangen, Germany), using a protocol as previously described.

The cranial MRI scans of the 32 patients were re-reported, mixed in with scans from patients matched for CD4 lymphocyte count who had also presented with neurological symptoms and signs and whose MR scans showed focal lesions, by two radiologists (MHC and BK) who were experienced in interpreting cranial MRI of patients with HIV. Scans were reported independently by the two radiologists: when there was disagreement consensus was reached in conference. The radiologists were blinded to the subject’s clinical and pathological diagnoses, and also to who was included in the study. Predetermined criteria were used for the MRI; in particular the number and anatomical location of lesions, their size, the degree of surrounding oedema, and the presence and nature of any enhancement was noted.

SPET of the brain was performed 30 minutes after injection of 3 mCi (111 MBq) of 201Tl intravenously. Patients were studied with one of two brain imaging instruments, either the SME (Strichman Medical Equipment) 810 brain dedicated tomograph (19 patients) or the GE (General Electric) Optima (13 patients). Data acquisition and reconstruction followed standard protocols. For the SME slice thickness was 1.25 cm and reconstruction was performed using an iterative method using filters set automatically by the manufacturer. For the Optima slice thickness was 2 pixels = 8 mm and prefiltering was performed using a Hann filter with a frequency of 0.8 cycles/cm, followed by standard ramp projection filter. Attenuation correction was performed using manufacturer’s algorithms. All SPET images were interpreted by an experienced nuclear medicine physician (DCC) who was “blinded” to the patient’s presumptive diagnosis. An initial qualitative (visual) assessment was carried out directly from the computer display.

In each patient where focal accumulation of 201Tl was seen anatomical correlation with focal lesions on MRI was performed and quantitative analysis was carried out using transaxial SPET images. For the SME images (SME810) or circular (SME 810) regions of interest (ROI) were used to obtain average radioactivity counts in each of the focal accumulations of 201Tl and in normal brain tissue. For background/normal brain tissue five ROI per slice (minimum of five slices) were used; sampling was done from areas within the same regions in the contralateral brain, unless there was a further lesion in that site, in which case other unaffected regions were selected. Background ROI were not chosen by selecting the lowest areas of contralateral uptake and particular care was taken to avoid highly vascular structures—that is, the scalp, meninges, and vessels in the base of the brain and the (sagittal and transverse) sinuses. The size of the ROI of both methods was consistent for both lesions and background. An uptake ratio was calculated for each 201Tl SPET focal abnormality from the ratio of activity in the lesion to the averaged background/normal brain tissue activity.

Lumbar puncture was performed in those patients without a contraindication (such as mass lesion or midline shift present on MRI). In addition to routine biochemical, microbiological, and cytological analyses tests for antibodies to T gondii were carried out (as described above). An aliquot of CSF was analysed at the same time for the presence of EBV, CMV, varicella zoster virus (VZV), herpes simplex virus type 1 (HSV-1), and type 2 (HSV-2) and JC virus using nested PCR amplification with oligonucleotide primers to detect EBV internal repeat (EBV-1), EBV gD, HSV-2 gG, and JC virus T antigen, as previously described.
Six patients had CT guided stereotactic brain biopsy and six had necropsy (two had both). All necropsies were performed by SBL. Neuropathological examination of necropsy tissue was performed by SBL and FS and brain biopsy tissue by FS. A protocol as previously described was used.

In addition, tissue was specifically examined for evidence of (a) lymphoma—identified by the presence of aggregations of pleomorphic lymphocytes either diffusely infiltrating the brain tissue or angiocentric in distribution. In addition, immunocytochemistry was performed using the B lymphocyte marker L26 (1:250; Dako Ltd, High Wycombe) and UCLH-1 (T lymphocyte marker; 1:250; Dako Ltd, High Wycombe). All lymphomas were L26 positive; (b) toxoplasmosis—identified by necrotising abscess formation with infiltration of inflammatory cells and macrophages with the presence of T gondii organisms on routine haematoxylin and eosin stain and/or immunohistochemical staining with polyclonal antibodies for T.gondii (1:150; ICN Pharmaceuticals, Thame).

The following definitions were used for diagnosis:

Toxoplasmosis, probable: typical cranial MRI appearances of single or multiple focal lesions with gadolinium enhancement and/or surrounding vasogenic oedema with partial or complete response to anti-toxoplasmosis therapy (without use of corticosteroids) seen on followup MRI.

Toxoplasmosis, definite: as above, with histological confirmation by necropsy.

Lymphoma, probable: MRI appearances (as above) failing to respond to anti-toxoplasmosis therapy, with or without detectable EBV DNA in CSF.

Table 1: Clinical and pathological diagnoses, CSF findings, MRI and SPET abnormalities

<table>
<thead>
<tr>
<th>Patient No</th>
<th>Co-trimoxazole prophylaxis</th>
<th>Blood T gondii serology (titre)</th>
<th>T gondii serology</th>
<th>EBV DNA</th>
<th>No of lesions</th>
<th>Site</th>
<th>Site(s) of accumulation</th>
<th>Uptake ratio(s)</th>
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<tbody>
<tr>
<td>Lymphoma, definite</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>No</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>4</td>
<td>(L)+(R) basal ganglia (perivenricular)</td>
<td>Periventricular, frontal white matter, (L)+(R) temporal lobes, brain stem</td>
<td>3.7–4.8</td>
</tr>
<tr>
<td>2</td>
<td>No</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>1</td>
<td>(L) temporal lobe</td>
<td>(R) parietal-occipital</td>
<td>2.1</td>
</tr>
<tr>
<td>3</td>
<td>Yes</td>
<td>–</td>
<td>Not done</td>
<td>1</td>
<td>(R) thalamus and septum paliucidum</td>
<td>(R) hemisphere midline</td>
<td>3.3</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Yes</td>
<td>+ (1:1024)</td>
<td>Not done</td>
<td>1</td>
<td>(R) fronto parietal</td>
<td>(R) frontoparietal</td>
<td>4.0</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>No</td>
<td>–</td>
<td>Not done</td>
<td>1</td>
<td>(L) temporal</td>
<td>(L) temporal</td>
<td>6.6</td>
<td></td>
</tr>
<tr>
<td>6</td>
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<td>–</td>
<td>Not done</td>
<td>2</td>
<td>(R) occipital and (R) temporal</td>
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<td>7</td>
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<td>(R) parietal</td>
<td>7.0</td>
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<tr>
<td>8</td>
<td>No</td>
<td>–</td>
<td>Not done</td>
<td>2</td>
<td>(L) basal ganglia</td>
<td>(L) perimedial hemisphere</td>
<td>2.2</td>
<td></td>
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<td>–</td>
<td>Not done</td>
<td>4</td>
<td>(R) thalamus, (L) basal ganglia</td>
<td>(R) deep frontal and parietal</td>
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<td></td>
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<td>Lymphoma, probable</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>Yes</td>
<td>–</td>
<td>Not done</td>
<td>4</td>
<td>(L)+(R) frontal, (R) temporal</td>
<td>(L) frontal</td>
<td>2.6</td>
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<tr>
<td>11</td>
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<td>–</td>
<td>Not done</td>
<td>10</td>
<td>(L) fronto parietal, (L) temporal</td>
<td>(L)+(R) basal ganglia</td>
<td>3.4</td>
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<td>–</td>
<td>Not done</td>
<td>2</td>
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<td>(L) occipital</td>
<td>3.1</td>
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<td>2</td>
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<td>(L) frontoparietal</td>
<td>3.0–2.1</td>
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<td>brain stem, periventricular (IV ventricle)</td>
<td>(L) posterior fossa</td>
<td>2.7</td>
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<tr>
<td>15</td>
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<td>–</td>
<td>+</td>
<td>1</td>
<td>(R) basal ganglia</td>
<td>Multiple, both hemispheres</td>
<td>2.6</td>
<td></td>
</tr>
<tr>
<td>16</td>
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<td>–</td>
<td>Not done</td>
<td>10</td>
<td>(R) frontal, (L)+(R) basal ganglia</td>
<td>Multiple, both hemispheres</td>
<td>1.9</td>
<td></td>
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<td>–</td>
<td>–</td>
<td>+</td>
<td>3</td>
<td>(R) frontal, (R) basal ganglia brain stem (subependymal IV ventricle)</td>
<td>(R) posteroventricle and parieto-occipital</td>
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<td>–</td>
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<td>–</td>
<td>+</td>
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<td>(L) parietal</td>
<td>Multiple scattered</td>
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<td>Lymphoma + Toxoplasma, definite</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
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<tr>
<td>19</td>
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<td>&gt;14</td>
<td>(R) occipital</td>
<td>(R) occipital</td>
<td>2.8</td>
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<tr>
<td>20</td>
<td>No</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>2</td>
<td>(R) temporal, (R) basal ganglia</td>
<td>(R) Frontal</td>
</tr>
<tr>
<td>Toxoplasmosis, probable</td>
<td></td>
<td></td>
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<td></td>
<td></td>
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<td>–</td>
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<tr>
<td>22</td>
<td>No</td>
<td>–</td>
<td>Not done</td>
<td>10</td>
<td>(R) frontal, (L) temporal brain stem</td>
<td>(R) hemisphere (&gt;3)</td>
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<tr>
<td>23</td>
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<td>–</td>
<td>Not done</td>
<td>10</td>
<td>(L)frONTAL, (L) TEMPORAL, (R) occipital</td>
<td>(L) posterior fronto</td>
<td>1.9</td>
<td></td>
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<tr>
<td>24</td>
<td>No</td>
<td>–</td>
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<td>2</td>
<td>(L)+(R) basal ganglia</td>
<td>(R) frontoparietal</td>
<td>1.7</td>
<td></td>
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<tr>
<td>25</td>
<td>Yes</td>
<td>–</td>
<td>Not done</td>
<td>10</td>
<td>periventricular (IV, III, and IV ventricles)</td>
<td>(L) posterior fossa</td>
<td>1.5</td>
<td></td>
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<tr>
<td>26</td>
<td>Yes</td>
<td>+ (1:250)</td>
<td>Not done</td>
<td>1</td>
<td>(L) frontal</td>
<td>Multiple, both hemispheres</td>
<td>1.4</td>
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<tr>
<td>27</td>
<td>No</td>
<td>–</td>
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<td>10</td>
<td>(L)+(R) basal ganglia</td>
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<td>28</td>
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<td>3</td>
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<td>None</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>29</td>
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<td>–</td>
<td>Not done</td>
<td>3</td>
<td>(L) frontal, (L) temporal, (L)+(R) basal ganglia</td>
<td>None</td>
<td>0</td>
<td></td>
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<tr>
<td>30</td>
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<td>+ (1:1024)</td>
<td>Not done</td>
<td>10</td>
<td>(R) frontal, (R) temporal (R) cerebellar hemisphere</td>
<td>(L)+(R) basal ganglia brain stem (periventricular)</td>
<td>None</td>
<td>0</td>
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<tr>
<td>31</td>
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<td>+ (1:1024)</td>
<td>–</td>
<td>–</td>
<td>5–10</td>
<td>(L) frontal, (R) occipital (L) cerebellar hemisphere</td>
<td>(R) basal ganglia</td>
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</tr>
<tr>
<td>32</td>
<td>No</td>
<td>–</td>
<td>Not done</td>
<td>5–10</td>
<td>(R) parietal, (L) temporal</td>
<td>(L)+(R) basal ganglia (subependymal around lateral ventricles)</td>
<td>None</td>
<td>0</td>
</tr>
</tbody>
</table>

+= positive; − = negative; EBV = Epstein–Barr virus; L = left; R = right.
Lymphoma, definite: as above, with histological confirmation by brain biopsy and/or necropsy.

Results of MRI and $^{201}\text{Tl}$ SPET were compared using a two tailed Fisher’s exact test. In those with toxoplasmosis the possible effect of duration of anti-toxoplasma therapy on $^{201}\text{Tl}$ SPET accumulation was assessed by the Mann-Whitney U test. A p value of <0.05 was considered significant.

Results

There were nine cases of definite lymphoma, nine cases of probable lymphoma, and 12 cases of probable toxoplasmosis. Two further cases had definite toxoplasmosis and lymphoma. In those with probable lymphoma, while the diagnosis was not confirmed histologically, the MRI appearances failed to respond to anti-toxoplasmatic therapy and were atypical for cryptococcosis, which do not enhance with gado- toxoplasmatic antibody IgM or with CSF analysis. The study population consisted of 23 patients. MRI of 23 patients showed multiple lesions (table 1), in the remainder a single lesion was demonstrated. Eight patients with lymphoma and one patient with toxoplasmosis had single lesions, p = 0.044, two tailed Fisher’s exact test. The interval between cranial MRI and $^{201}\text{Tl}$ SPET scanning ranged between 1 and 25 (median 7) days.

SPET scanning showed focal accumulation of $^{201}\text{Tl}$ in 24 patients. No significant difference was noted between the two cameras for detection of uptake in deep lesions. Only one of 18 lymphoma cases had no focal accumulation while six of the 12 with toxoplasmosis showed no uptake, p = 0.034, two tailed Fisher’s exact test. One patient with both lymphoma and toxoplasmosis had no focal accumulation of $^{201}\text{Tl}$. In all other patients, uptake ratios varied widely (table 1). In those with lymphoma (including the two patients with both lymphoma and toxoplasmosis) uptake ratios ranged from 0 to 7.6 (median 2.85) and of those with toxoplasmosis uptake ratios ranged from 0 to 2.1 (median 0.7). Of the 18 patients with lymphoma, six patients had one or more lesions seen on MRI which were ≤18 mm and did not accumulate $^{201}\text{Tl}$ (table 1). If the criteria used by O’Malley et al. are applied to these data and an uptake ratio of ≥2.9 is taken to represent lymphoma, then this finding has a sensitivity of 55%, a specificity of 100%, and a negative predictive value of 57% for diagnosis of lymphoma. In addition, if an uptake ratio of ≤2.0 is used to determine which patients have toxoplasmosis, this finding has a sensitivity of 83%, a specificity of 75%, and a negative predictive value of 88%. Of eight patients with single lesions on MRI and focal accumulation of $^{201}\text{Tl}$ only one proved to be toxoplasmosis.

In all but two patients (Nos 2 and 6) anti-toxoplasma therapy had been commenced at the time of SPET scanning, median 7 days (range 1–42 days). In those patients with toxoplasmosis, if those who had received ≤7 days anti-toxoplasma therapy are compared with those who had received >7 days treatment, no differences in $^{201}\text{Tl}$ SPET ratios were observed: Mann–Whitney U test (p=NS).

Discussion

This study has sought to define the accuracy with which focal CNS lesions due to lymphoma can be distinguished from toxoplasmosis by neuroimaging using MRI and $^{201}\text{Tl}$ SPET and laboratory investigations including CSF analysis. The study population consisted of a selected series of patients infected with...
HIV and well documented neurological presentations. The results show that the MRI findings of single or multiple focal lesions can represent either lymphoma or toxoplasmosis, with both kinds of lesions showing ring or other patterns of enhancement. This lack of discrimination using MRI has previously been reported and our observations are confirmatory. However certain specific MRI abnormalities in our study were strongly suggestive of lymphoma—for example, if lesions were single or were periventricular in location and demonstrated subependymal spread. These clinically useful appearances were only present in a minority however.

In our study seven of 32 patients had positive serology for toxoplasmosis. Eight of the 14 patients with toxoplasmosis had no detectable IgG antibodies and, in contrast, two with histologically confirmed lymphoma and no toxoplasmosis had positive serology. Thus we were unable to use toxoplasma serology to aid discrimination between diagnoses. Similarly we found that knowledge of patients’ use of prophylaxis with co-trimoxazole did not aid discrimination. These data contrast with those from a recent study of 97 HIV infected patients from Rome11 where toxoplasma serology was positive in 97% of those with toxoplasmosis and in only 3.1% with lymphoma. Using a decision making analysis model and applying Bayes’ theorem, it was found in those patients with positive toxoplasma serology who were not taking anti-P. carinii prophylaxis which was also effective against toxoplasmosis, the probability of toxoplasmosis was 0.87, but was only 0.59 if prophylaxis was given. In those with negative toxoplasma antibodies the probability of lymphoma was 0.74. Several other studies have suggested that, in the HIV infected population, possession of IgG antibodies is strongly associated with the development of toxoplasmosis. Interpretation of these data is confounded by widely varying rates of seroprevalence in the general population in different geographical locations. In France approximately 75% of adults have antibodies where as in the United States and the United Kingdom the rate is approximately 25%

In those patients in our study, in whom lumbar puncture was carried out, EBV DNA detection was strongly associated with CNS lymphoma. However we were only able to perform CSF analysis in 8/32 (25%) of our patients. In the study from Rome11 CSF was obtained in only 38/97 (40%) of patients. In that study, if EBV DNA was detected in CSF then the probability of lymphoma was 0.96. These data and those from our own study underscore the fact that although genome detection from CSF is a highly sensitive and specific technique for diagnosis, any such molecular diagnostic strategy is limited by the inability to perform lumbar puncture in a proportion of patients with focal CNS lesions because of contraindications such as mass effect.

In our study we found focal accumulation of 201Tl in all but one of the 18 patients with lymphoma, and in one of the two patients with toxoplasmosis. In contrast, accumulation was seen in only half of those with toxoplasmosis. High uptake ratios, >2.9, were only found in those with lymphoma. However, some patients with lymphoma had lower uptake ratios, between 1.5 and 2.1, as did patients with toxoplasmosis. These data contrast with a previous study which suggested that measurements of the uptake ratios in those with focal accumulation of 201Tl aided discrimination between lymphoma and toxoplasmosis. In this study, from Washington, of 13 HIV infected patients with focal CNS lesions on MRI six patients had 201Tl uptake ratios (calculated using a method similar to ours) of between 2.95 and 4.30 (mean 3.65): all these patients had lymphoma. Seven patients had uptake ratios ranging from 0.77 to 1.95, five had toxoplasmosis (one also had CMV and Mycobacterium tuberculosis), one had progressive multifocal leuкоencephalopathy, and one had a venous angioma.

In a prospective study from Miami 37 HIV infected patients with focal CNS lesions on cranial CT or MRI also had imaging with 201Tl SPET.18 Intense focal accumulation was seen only in 12 patients, all of whom had biopsy or necropsy confirmed lymphoma. No accumulation of 201Tl was seen in the other 25 patients, of whom 15 had toxoplasmosis. In a third study from New York, 18 HIV infected patients with focal CNS lesions underwent 201Tl SPET imaging.19 In this study 10 patients had focal accumulation of 201Tl in focal CNS lesions shown by CT/MRI with uptake ratios of between 2.1 and 30.6. Of these five had biopsy proved lymphoma, three had probable lymphoma (using criteria similar to those used in our study), one had metastatic adenocarcinoma, and one had probable toxoplasmosis. Eight other patients had no accumulation of 201Tl and these seven had toxoplasmosis (six by clinical response, one confirmed by biopsy) and one had a cryptococcomaa.

These three studies differed from ours in several ways. Firstly, 201Tl SPET was performed in the majority of patients within 72 hours of CT or MRI. Secondly, the imaging protocol in two of the studies used higher doses of radiotracer and imaged more rapidly after intravenous injection of 201Tl. Imaging was 5 minutes after 5 mCi 201Tl in one study18 and 10 minutes after 4 mCi in the other.19 However, the study18 which showed 201Tl SPET to clearly discriminate between lymphoma and toxoplasmosis used an identical imaging protocol to ours. The third difference lies in the patients themselves. In our study of eight patients with single lesions on MRI and focal accumulation of 201Tl seven had lymphoma. These seven patients all had uptake ratios >2.0 which enabled discrimination from the patient with toxoplasmosis to be made as in this case the uptake ratio was only 1.4. In the study from Washington17 all 13 patients had only single focal CNS lesions on MRI. In the study from Miami18 of the 37 patients 13 had a single mass lesion. Accumulation was seen only in the seven with lymphoma and not in the six
with toxoplasmosis. Eight of the 18 patients in the study from New York had single mass lesions; focal accumulation occurred in all three with lymphoma as the cause, one of three with toxoplasmosis, and one patient with adenocarcinoma.

Other nuclear medicine techniques have been used to aid discrimination between toxoplasmosis and lymphoma including technetium-99m Sestamibi (\(\text{Tc}^9\text{m MIBI}\)) SPET, and fluorine-18 fluorodeoxyglucose positron emission tomography (\(\text{FDG PET}\)). SPET with \(\text{Tc}^9\text{m MIBI}\) was compared with \(\text{TI}^{201}\) in a study of 17 HIV infected patients with focal CNS lesions (four had biopsy proved lymphoma). For diagnosis of lymphoma both techniques had 100% sensitivity but a specificity and negative predictive value of 69% and 54% for \(\text{Tc}^9\text{m MIBI}\) and 54% and 40% for \(\text{TI}^{201}\). The higher preferential uptake by the choroid plexus of \(\text{Tc}^9\text{m MIBI}\) compared with \(\text{TI}^{201}\) means that in HIV infected patients it is more difficult to evaluate focal CNS lesions which are paraventricular in location as may occur in cerebral lymphoma. FDG PET which can measure regional metabolic activity accurately discriminates between toxoplasmosis and lymphoma (and other conditions including progressive multifocal leuco-encephalopathy). The disadvantage of this technique is that it is not available in the majority of centres and it is more expensive than \(\text{TI}^{201}\) SPET scanning.

In conclusion, in our study knowledge of patients’ toxoplasma serology and use of co-trimoxazole for PCP did not aid discrimination between lymphoma and toxoplasmosis. Detection of EBV DNA in CSF was strongly sensitive and specific for toxoplasma therapy is indicated. In those who fail to respond stereotactic brain biopsy remains an option. However, factors including the underlying condition of the patient, accessibility of lesions to biopsy, and decisions about which lesion to biopsy, together with the morbidity and mortality associated with the procedure should be considered carefully.

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Conflict of interests: None.

Adapted from the description of the MRI appearances of the patients in this study is held on file in the STI office and is available to interested readers.

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Magnetic resonance imaging, thallium-201 SPET scanning, and laboratory analyses for discrimination of cerebral lymphoma and toxoplasmosis in AIDS.

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