Interpretation of Chlamydia trachomatis antibody response in chlamydial oculogenital infection

H C Patel, B T Goh, N D Viswalingam, J D Treharne

Abstract

Objective—To study: (a) the chlamydial antibody response to the D-K serovars using the micro-immunofluorescence (micro-IF) test in the following groups: (I) chlamydial genital infection only, (II) chlamydial ocular infection only, (III) combined chlamydial ocular and genital infection (oculo-genital infection), (IV) chlamydial ocular infection with chlamydial-negative non-gonococcal urethritis, (V) adenosivirus conjunctivitis (control group 1), (VI) male partners of group I-IV with no chlamydial ocuogenital infection or non-gonococcal urethritis (control group 2) (b) the cross reactivity of antibodies in patients’ sera between the three chlamydial species and within the serovars of C trachomatis in those with culture-positive chlamydial oculo-genital infection.

Setting—oculogenital (diagnostic) clinic at Moorfields Eye Hospital, London, UK.

Subjects—209 consecutive patients attending the clinic with Chlamydia trachomatis oculogenital infection and 86 patients with adenovirus conjunctivitis (control group 1) and 55 male partners with no evidence of chlamydial oculo-genital infection or non-gonococcal urethritis (control group 2).

Results—Of all the patients with proven chlamydial ocuogenital infection, 10-5% (22/209) and 94% (197/209) had IgM and IgG antibodies respectively. The geometric mean IgG antibody titres (GMT) were 1:98, 1:123, 1:245 and 1:101 in groups I to IV respectively. The IgG GMT values seen in control groups 1 and 2 were 1:45 and 1:36 respectively. Only 2/86(2%) patients in group V (control group 1) had IgG chlamydial antibodies of 1:32 and 1:64, whilst only 1/55(1-8%) and 4/55(7-3%) of patients in group VI(control group 2) had chlamydial IgG antibody titres of ≥1:256 and ≥1:128 respectively. A four-fold rise or fall in IgG antibody titre occurred in 56%(107/192) of patient groups I-IV over 2-6 weeks. Low titre cross-reactive antibody responses against different chlamydial species and serovars were commonly seen; 71%(148/209) of all patients showed cross-reactivity with Chlamydia pneumoniae or psittaci species or both, whilst 92% (193/209) of patients showed some level of cross-reactivity to other pooled serovars of C trachomatis (A-C and L 1-3).

Conclusions—Serological diagnosis of chlamydial infection as evidenced by a positive IgM antibody response, high IgG titre (≥1:256) or ≥4-fold rise or fall in IgG antibody titre was seen in 78%(163/209) of patients with culture-positive chlamydial oculo-genital infection. Chlamydial IgG antibody titres of ≥1:256 had a sensitivity of 42.6%, specificity of 98.2%, positive predictive value of 98.8% and a negative predictive value of 31% for chlamydial infection at any site, when considering groups I-IV and control group 2. In this study of 216 patients with conjunctivitis, a positive IgG antibody response (titre ≥1:16) had a sensitivity of 98.5%, specificity of 97.7%, positive predictive value of 98.5% and a negative predictive value of 97.7%, for chlamydial conjunctivitis. Patients with dual chlamydial infection of conjunctiva and genital tract had a higher IgG GMT titre than those with ocular or genital infection alone: infection at a second site may produce an anamnestic response. Although the micro-IF test is a useful adjunct for the diagnosis of chlamydial infection, cross-reactivity between different chlamydial species and serovars is common. Chlamydial seroepidemiological studies should be interpreted with caution, as studies may attribute a serological response to a particular species or serovar in a setting where two or more are prevalent.

Introduction

Three major problems have precluded the successful use of serological tests for the diagnosis of an individual patient’s chlamydial genital infection. First, the baseline prevalence of antibody in populations which are sexually active and likely to be at risk for Chlamydia trachomatis infection is high, often ranging from 45 to 65% of persons tested. Also little is known about the predictive value of a single antibody test.

The second major difficulty precluding effective use of serodiagnosis results from the asymptomatic or minimally symptomatic nature of many chlamydial infections. Lack of an abrupt onset of symptoms means that many patients are seen during periods when IgM antibody or rising or falling titres of IgG antibody cannot be demonstrated.
The third major problem is the cross-reactivity between the different species of chlamydia as well as between different serovars of *C. trachomatis*. This makes it difficult to interpret an antibody response to a particular species in a setting where two or more chlamydial species may be prevalent, for example *C. trachomatis* and *C. pneumoniae*, and also this also questions the validity of studies attributing various clinical syndromes to *C. trachomatis* or other species based on serological evidence alone.

Early serological tests for the diagnosis of chlamydial infection included the complement fixation test (CFT) which may be useful for the diagnosis of the systemic chlamydial infections, lympho-granuloma venereum and psittacosis. However the CFT only detects antibody to the heat-stable, lipopolysaccharide antigen common to all *chlamydia* species and cannot distinguish between infections due to *C. trachomatis*, *C. psittaci* or *C. pneumoniae*. In 1970, Wang and Grayston described an indirect immunofluorescence serotyping test (micro-IF), which has led to the elucidation of 15 different serovars of *C. trachomatis*, and has now become accepted as the serological standard. The detection of type-specific antibodies using all 15 serovar antigens can be time-consuming and expensive and modifications of the original test have been described using pooled serologically related antigens, or pooled, epidemiologically related serovars. In sera from patients with chlamydial infections, broadly cross-reactive antibody responses to different species and serovars of chlamydia may commonly occur. We were interested to study the micro-IF chlamydial antibody response to the D-K and cross-reactivity between the A-C, D-K and L 1-3 serovars as well as between the different species of chlamydia, seen in patients with culture proven chlamydial ocular-genital infection. In particular, we wanted to ascertain the role of the serological test in diagnosing chlamydial ocular-genital infection.

### Methods

Between April 1987 and February 1992, 209 consecutive patients attending the Diagnostic Clinic at Moorfields Eye Hospital with chlamydial ocular-genital infection were retrospectively analysed. Cultures for *Chlamydia trachomatis* were performed from the conjunctiveae of patients with conjunctivitis. Cultures were also performed from the cervix, urethra and rectum in women and urethra of heterosexual men who had or whose partners had chlamydial conjunctivitis. Screening for other genital infections was also performed. All men were screened for non-gonococcal urethritis (≥10 pus cells/HPF × 1000 magnification in the urethral smear). According to the clinical findings and chlamydial culture results, the patients were categorised as above.

Serology was performed in all patients for syphilis and chlamydial IgG and IgM antibodies. A modified microimmunofluorescence test using pooled antigens was used to detect chlamydial antibodies.

### Results

Of all patients with proven chlamydial ocular-genital infection 10-5% (22/209) and 94% (197/209) had positive IgM (≥1:8) and IgG (≥1:16) antibody titres to the D-K serovars respectively, at initial examination (table 1). However, one IgM negative patient had a positive IgM chlamydial antibody titre and seven patients IgG negative at outset subsequently seroconverted on serial testing, leaving only four (2%) patients with a persistent negative IgG response to chlamydia.

The geometric mean antibody IgG titres (GMT) were 1:98, 1:123, 1:245 and 1:101 in groups I to IV respectively. The IgG GMT values in control groups 1 and 2 were 1:45 and 1:36 respectively. Two of 86 (2%) patients in group V (control group 1) who were of a similar age range had IgG antibody titres of 1:32 and 1:64. Only 1/55 (1.8%) and 4/55 (7.3%) of patients in group VI (control group 2) had chlamydial IgG antibody titres of ≥1:256 and ≥1:128 respectively. No IgM response was seen in the control groups. In 192 patients who had serial serum specimens taken, IgG chlamydial antibody titres in blood rose by 4-fold or more in 38 (20%) patients within the first six weeks following presentation and in an additional 69 (36%) patients antibody titres fell by 4-fold or more, during the same period. Thus in 107 out of 192 (56%) patients a classical serodiagnostic response was shown (table 2).

Some low-titred cross-reactive IgG antibody response was seen in the majority of patients. Cross-reactivity between chlamydial species is shown in the Venn diagram (fig).

In the majority, (802/836(96%)) of the serological tests that showed cross-reactivity, the IgG antibody titres to other *C. trachomatis* serovars or chlamydial species were either equal (224/836)(26-8%) or lower or negative (578/836(69%) than were the antibody titres to the D-K serovars (table 3).

### Table 1 IgM and IgG response in chlamydial ocular-genital infection

<table>
<thead>
<tr>
<th>Infection Site</th>
<th>Total</th>
<th>IgM</th>
<th>IgG</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td>Genital</td>
<td>79 (100%)</td>
<td>70 (87%)</td>
<td>9 (13%)</td>
</tr>
<tr>
<td>Ocular</td>
<td>20 (100%)</td>
<td>16 (75%)</td>
<td>4 (25%)</td>
</tr>
<tr>
<td>Oculogenital</td>
<td>80 (100%)</td>
<td>73 (90%)</td>
<td>7 (10%)</td>
</tr>
<tr>
<td>Ocular &amp; NSU</td>
<td>30 (100%)</td>
<td>28 (93%)</td>
<td>2 (7%)</td>
</tr>
<tr>
<td>Total</td>
<td>209 (100%)</td>
<td>187 (89%)</td>
<td>22 (10-5%)</td>
</tr>
</tbody>
</table>

* Titre ≥ 1:16
* 2 had subsequent IgM sb
* 4 had subsequent IgM ab
* 1 had subsequent IgM sb

### Table 2 Chlamydia serial IgG antibody response in ocular-genital infection

<table>
<thead>
<tr>
<th>Site of Infection</th>
<th>No Change</th>
<th>≥ 4 fold f</th>
<th>≥ 4 fold d</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ocular</td>
<td>6 (35%)</td>
<td>8 (47%)</td>
<td>3 (18%)</td>
<td>17 (100%)</td>
</tr>
<tr>
<td>Genital</td>
<td>40 (55%)</td>
<td>13 (18%)</td>
<td>20 (27%)</td>
<td>73 (100%)</td>
</tr>
<tr>
<td>Oculo-Genital</td>
<td>30 (41%)</td>
<td>7 (9%)</td>
<td>37 (50%)</td>
<td>74 (100%)</td>
</tr>
<tr>
<td>Ocular &amp; NSU</td>
<td>9 (32%)</td>
<td>10 (36%)</td>
<td>9 (32%)</td>
<td>28 (100%)</td>
</tr>
<tr>
<td>Total</td>
<td>85 (44%)</td>
<td>38 (20%)</td>
<td>69 (36%)</td>
<td>192 (100%)</td>
</tr>
</tbody>
</table>
Table 3  Cross-reactive IgG antibody titres in C trachomatis oculo-genital infection

<table>
<thead>
<tr>
<th>Title*</th>
<th>C pneumoniae</th>
<th>C psittaci</th>
<th>C trachomatis (A-C)</th>
<th>C trachomatis (L 1-3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Higher</td>
<td>15 (7%)</td>
<td>2 (1%)</td>
<td>10 (5%)</td>
<td></td>
</tr>
<tr>
<td>Same</td>
<td>17 (8%)</td>
<td>8 (4%)</td>
<td>86 (41%)</td>
<td>113 (54%)</td>
</tr>
<tr>
<td>Lower</td>
<td>102 (49%)</td>
<td>89 (42%)</td>
<td>98 (47%)</td>
<td>68 (32%)</td>
</tr>
<tr>
<td>Negative</td>
<td>75 (36%)</td>
<td>110 (53%)</td>
<td>18 (9%)</td>
<td>18 (9%)</td>
</tr>
<tr>
<td>Total</td>
<td>209 (100%)</td>
<td>209 (100%)</td>
<td>209 (100%)</td>
<td>209 (100%)</td>
</tr>
</tbody>
</table>

*titre compared with C trachomatis (D-K) titre

Discussion

Confirmation by culture or antigen detection with enzyme immunosay (EIA), or direct immunofluorescence (DIF) are the preferred methods for diagnosing C trachomatis infections. However, in many cases this is not possible because certain sites such as Fallopian tubes or lungs are difficult to sample and some patients have had prior anti-chlamydial antibodies. In these cases, serology may be useful.

In this study the sensitivity, specificity, positive predictive value and negative predictive value for C trachomatis infection at any site was 42-6%, 98-2%, 98-8% and 31% respectively if the IgG chlamydial antibody titre was ≥1:256, when comparing groups I-IV with group VI as the negative control. Thus an IgG response of ≥1:256 is indicative of chlamydial infection in this group of patients.

IgM chlamydial antibody response was seen in the first serum specimens of only 10-5% (22/209) of patients with culture proven chlamydial oculo-genital infection, whereas 25% of patients with ocular infection alone, showed an IgM response compared with 7-13% in those with genital infection with or without ocular involvement. The lower response in the latter groups may be due to asymptomatic or minimally symptomatic chlamydial genital infection that was present in most of our patients for some time before presenting with ocular infection. The transient IgM response seen in early chlamydial genital infection has often disappeared by this time.5

A positive chlamydial IgG antibody titre of ≥1:16 and ≥1:64 was seen in the first serum of 94% (197/209) and 77% (161/209) of all our patients with culture proven chlamydial infection respectively. Similar antibody responses are seen in patients with chlamydial ocular infection6 and pelvic inflammatory disease.7 Treharne et al observed that 23% (83/143) and 62% (33/142) of women with acute salpingitis had a positive IgM (≥1:8) and IgG (≥1:64) response respectively.7

In this study 98-5% (128/130) of patients who had a chlamydia-positive ocular infection, with or without genital infection, had a positive serum IgG titre (≥1:16) to the D-K serovars at initial examination compared with 2/86 (2%) patients in the adenosivus conjunctivitis group. A positive chlamydial antibody IgG response in serum, in a patient with conjunctivitis in this study had a sensitivity, specificity, positive predictive value and negative predictive value of 98-5%, 97-7%, 98-5% and 97-7% respectively for chlamydial ocular infection. Darougar et al showed that the presence of antichlamydial IgG at a titre of ≥1:32 or IgM at a titre of ≥1:8 was closely associated with ocular paratrachoma while only 3% of 207 patients with non-chlamydial conjunctivitis had chlamydial antibodies in the serum.8

In individual patients the presence of high serum IgG titres (≥1:256), a four-fold or greater change in IgG titre, or the presence of serum IgM antibody may indicate chlamydial infection. In the highly selected group with conjunctivitis, chlamydial IgG antibody is a good indicator of chlamydial infection even at a low titre of ≥1:16. Serology therefore could be particularly useful in complicated cases such as children with chlamydial pneumonia8, or in patients partially treated with antibiotics.

Two percent (4/192) of patients were consistently negative for chlamydial IgG antibody on serial testing, although all these patients were found to have chlamydial genital infection by isolation, three were male and one was female. This level of serological non-reactivity is lower than other studies which showed that 10-20% of men with chlamydial urethral infection develop no detectable specific antibodies at all.** In men, the absence of the
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antibody response may be because of weak antigenic stimulus at a very localised mucosal site.

The GMT of the antibody response in chlamydial ocular genital infection was lowest in those with primary genital infection, particularly in men, whilst it was highest in females who had dual ocular and genital chlamydial infection. This may be because of a greater antigenic stimulus and anamnestic response resulting from a second infected site, superimposed on long standing asymptomatic genital infection leading to a larger surface area of epithelium being involved. Cross reactive IgG antibody responses were seen between most chlamydial species and between the serovars of C trachomatis. The IgG titre was the same (26-8%), or lower or negative for chlamydial antibodies (69%) than to that of the D-K serovars in patients with chlamydial-positive ocular genital infection, probably indicating serum cross-reactivity rather than an active infection due to other serotypes in these patients. Kuritsky et al showed that of 55 patients who had ≥ 4 fold rise in titre to other respiratory pathogens, 3(5%) also had ≥ 4 fold rise in IgM or IgG antibodies to C trachomatis and seven of eight patients with psittacosis were also able to seroconvert to C trachomatis serovars. Our study showed cross-reactive IgG antibody to the D-K serovars in 96% (802/836) of sera tested.

The type-specific micro-IF test, and to a greater degree genus-specific tests such as CFT, single-antigen immunofluorescence or elementary-body based ELISA antibody tests for the detection of chlamydial infection, may also demonstrate cross-reaction to C pneumoniae. This organism is now a recognised cause of respiratory infection and may be associated with myocarditis, chronic coronary heart disease, acute myocardial infarction and sarcoidosis. Based on serological tests alone, C trachomatis has been reported to cause community acquired pneumonia and myocarditis in adults. Because of this, many of the previously reported genus-specific serological studies or those which excluded this chlamydial species need to be reassessed using specific C pneumoniae antigens. Although serovar and species-specific micro-IF tests are sensitive and can differentiate between species, they are technically difficult and only available in specialist laboratories. Cross reactions between chlamydial species and serovars even in micro-IF tests may give confusing results. Hence, it is desirable that cultures or chlamydial antigen detection methods such as EIA or DIF are performed to detect chlamydial infection and to assess the diagnostic significance of serologic tests. Antigenic cross-reactivity, polyclonal antibody stimulation or cross infection with a related strain, or coexisting chlamydial respiratory pathogens may be important confounding and confusing factors in interpreting serologic tests for the diagnosis of chlamydial ocular genital infections. In the event that it is impractical to perform chlamydial culture or to carry out chlamydial antigen detection tests in support of clinical diagnosis, then the micro-IF serological test may be useful but should be interpreted with caution.

12 Grayston JT, Kuo C, Wang SF, Altman J. A new Chlamydia psittaci strain, TWAR, isolated in acute respira-