Analysis of quinolone resistance mechanisms in Neisseria gonorrhoeae isolates in vitro

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Background and objectives: Gonococcal fluoroquinolone resistance is now a significant problem in Japan. We generated gonococcal mutants resistant to norfloxacin in vitro from norfloxacin sensitive isolates and analysed the contribution of three known mechanisms of quinolone resistance in Neisseria gonorrhoeae.

Materials and methods: Three clinical isolates of N gonorrhoeae susceptible to norfloxacin were exposed to increasing concentrations of norfloxacin. To identify mutations in the gyrA and parC genes of the gonococcal mutants, the quinolone resistance determining regions of the gyrA and parC genes were polymerase chain reaction (PCR) amplified and the PCR products were directly sequenced. Norfloxacin accumulation in the gonococcal cells was also measured.

Results: The MICs of norfloxacin for three variants containing a single GyrA mutation were 16-fold higher than that for their parent isolates. A variant showing reduced norfloxacin accumulation in the cells, without mutations in the GyrA or ParC proteins, was also less sensitive to norfloxacin, with a 16-fold increase in the MIC, compared with the parent strain. The MIC of norfloxacin for a variant which contained a single GyrA mutation with reduced norfloxacin accumulation in the cells was 128-fold higher than for the parent strain. A variant containing mutations in both GyrA and ParC proteins with reduced accumulation of norfloxacin in the cells showed a 256-fold increase in the norfloxacin MIC compared with the parent strain. There was no variant containing a ParC mutation without the simultaneous presence of a GyrA mutation.

Conclusions: The results from this study suggest that not only a mutation in the gyrA gene but also reduced drug accumulation in cells contributes to the development of fluoroquinolone resistance in gonococci and that a mutation in the parC gene with the simultaneous presence of a mutation in the gyrA gene contributes to a high level of fluoroquinolone resistance in gonococci with decreases in accumulation in cells having an additional but lesser effect.

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Keywords: quinolone resistance; Neisseria gonorrhoeae; mutation; gyrA gene; parC gene; norfloxacin accumulation

Introduction
Fluoroquinolones are very effective as oral single dose treatment for infections caused by Neisseria gonorrhoeae. Fluoroquinolone regimens have been increasingly used in various countries including Japan. Recently, gonococcal isolates with reduced susceptibility to fluoroquinolones have been identified in several countries. In Japan, gonococcal fluoroquinolone resistance is now a significant problem.

In N gonorrhoeae, three mechanisms of fluoroquinolone resistance have been reported: (i) the development of mutations in the DNA gyrase subunit A (GyrA) encoded by the gyrA gene, (ii) mutations in the DNA topoisomerase IV encoded by the parC gene, and (iii) reduced quinolone accumulation in the cells. Mutations at serine-91 and at aspartic acid-95 in the gonococcal GyrA protein are the commonly identified mutations in quinolone resistant isolates. However, no single mutation in the parC gene, without the coexistence of a mutation in the gyrA gene, has been detected. These findings suggest that in the development of gonococcal fluoroquinolone resistance a mutation within the gyrA gene may be more important than that within the parC gene. Reduced accumulation of quinolone within bacterial cells is also known to play a role in the quinolone resistance in several bacterial species. However, its mechanism remains unclear in several points in N gonorrhoeae.

We are interested in determining how a N gonorrhoeae isolate acquires quinolone resistance in vitro. We generated gonococcal variants resistant to norfloxacin by exposing three clinical isolates susceptible to norfloxacin to increasing concentrations of norfloxacin in vitro. We analysed the contributions of the three known mechanisms of quinolone resistance.

Materials and methods
BACTERIAL STRAINS
N gonorrhoeae wild type (WT) strains used in this study were A67/WT, A69/WT, and A219/WT. The three strains were isolated from Japanese men with acute urethritis. N gonorrhoeae was identified as Gram negative diplococcus on Gram stain and by the oxidase reaction and the sugar utilisation pattern.

SELECTION OF THE N GONORRHOEAE RESISTANT TO NORFLOXACIN
Each wild type strain was cultured overnight on a GC plate (Becton Dickinson, USA). Approximately 10 colony forming units (cfu) of
each strain were then plated on GC agar containing norfloxacin (1 × minimum inhibitory concentration (MIC)). These plates were incubated at 35°C for 48 hours in a 5% carbon dioxide atmosphere. Growing bacteria were replated on GC agar containing the same concentration of norfloxacin for one additional 48 hour cycle. In the following selection step, bacteria were cultured on GC agar containing a doubled concentration (2 × original MIC) of norfloxacin. A resistant isolate was grown up and replated as before on GC plates containing the same concentration of norfloxacin. This protocol was repeated for additional cycles. At each step, single colonies were collected for susceptibility testing, DNA sequencing, and an assay of norfloxacin accumulation in the cells.

ANTIBIOTIC SUSCEPTIBILITY TESTING
The MIC for the wild type and norfloxacin selected variants was determined using an agar dilution technique with a GC agar base containing 1% IsoVitaleX (Becton Dickinson) and twofold dilutions of the antibiotic. Plates were inoculated with 5 µl of 10⁶ cfu/ml of each isolate using a multipoint inoculator. The plates were incubated at 35°C for 24 hours in a 5% carbon dioxide atmosphere. MICs were defined as the lowest antibiotic concentration that inhibited bacterial growth. The antibiotics which were tested were three fluoroquinolones: norfloxacin, ciprofloxacin, and DU6859a, as well as ampicillin, ceftriaxone, cefotetam, imipenem, azithromycin, and spectinomycin. All of the antibiotics were obtained in powder form from their manufactures. β lactamase production was tested by an acidoemtric assay (β check; Pfizer Pharmaceuticals Inc, Japan).

EXTRACTION OF DNA AND ANALYSIS
Polymerase chain reaction (PCR) and direct DNA sequencing were performed to identify mutations in the gyrA and parC genes in the norfloxacin selected variants. Chromosomal DNA was extracted using standard methods and was subjected to PCR. The oligonucleotide primers which were used in the PCR amplification of the gyrA gene have been described previously. For amplification of the parC gene, two primers (the forward primer: 5'-ATGCCGCGATAGGTGTTGAC-3' and the reverse primer: 5'-AACCGCTTAACG ACAACAGG-3') were produced by a DNA synthesiser, according to the sequences which have been reported previously. The parC gene sequence was determined from nucleotides 166 to 420 which correspond to the amino acids from 56 to 140 of the gonococcal ParC protein. This includes the quinolone resistance determining region (QRDR) (amino acids 66 to 119 of the gonococcal ParC protein). The MICs of norfloxacin for isolates A67/A and A67/B were 16 and 256-fold higher than that for the parental isolate after being cycled 18, 14, and 10 times, respectively.

A significant reduction in the sensitivity to norfloxacin was observed in the six norfloxacin selected variants (table 1). The MICs of norfloxacin for isolates A67/A and A67/B were 16 and 256-fold higher than that for the parental isolate A67/WT. Variants A69/A and A69/B were also less sensitive to norfloxacin, with a 16 and 128-fold increase in the MICs, respectively, compared with the parent strain A69/WT. The MICs of norfloxacin for both A219/A and A219/B strains were 16-fold higher than for the parent strain A219/WT. The six norfloxacin resistant variants demonstrated cross-resist-
Table 1  Susceptibilities of wild type and variant Neisseria gonorrhoeae to various antibiotics

<table>
<thead>
<tr>
<th>Strain</th>
<th>NFLX (µg/ml)</th>
<th>CPFX</th>
<th>DU6859a</th>
<th>ABPC</th>
<th>CTRX</th>
<th>CFTM</th>
<th>IPM</th>
<th>AZM</th>
<th>SPCM</th>
</tr>
</thead>
<tbody>
<tr>
<td>A67/WT</td>
<td>0.016 (1x)†</td>
<td>0.004 (1x)</td>
<td>0.002 (1x)</td>
<td>0.25</td>
<td>0.031</td>
<td>0.063</td>
<td>0.063</td>
<td>0.016</td>
<td>4.0</td>
</tr>
<tr>
<td>A67/A</td>
<td>0.25 (16x)</td>
<td>0.063 (16x)</td>
<td>0.004 (2x)</td>
<td>0.25</td>
<td>0.031</td>
<td>0.125</td>
<td>0.125</td>
<td>0.016</td>
<td>4.0</td>
</tr>
<tr>
<td>A67/B</td>
<td>4.0 (256x)</td>
<td>0.5 (128x)</td>
<td>0.031 (1x)</td>
<td>0.25</td>
<td>0.031</td>
<td>0.063</td>
<td>0.063</td>
<td>2.0</td>
<td></td>
</tr>
<tr>
<td>A69/WT</td>
<td>0.004 (1x)</td>
<td>0.004 (1x)</td>
<td>0.002 (1x)</td>
<td>0.125</td>
<td>0.004</td>
<td>0.063</td>
<td>0.063</td>
<td>0.016</td>
<td>4.0</td>
</tr>
<tr>
<td>A69/A</td>
<td>0.063 (16x)</td>
<td>0.016 (4x)</td>
<td>0.002 (1x)</td>
<td>0.25</td>
<td>0.002</td>
<td>0.031</td>
<td>0.125</td>
<td>0.016</td>
<td>4.0</td>
</tr>
<tr>
<td>A69/B</td>
<td>0.008 (4x)</td>
<td>0.125</td>
<td>0.004 (1x)</td>
<td>0.125</td>
<td>0.004</td>
<td>0.16</td>
<td>0.063</td>
<td>0.31</td>
<td>8.0</td>
</tr>
<tr>
<td>A219/WT</td>
<td>0.031 (1x)</td>
<td>0.008 (1x)</td>
<td>0.025</td>
<td>0.004</td>
<td>0.16</td>
<td>0.063</td>
<td>0.031</td>
<td>8.0</td>
<td></td>
</tr>
<tr>
<td>A219/A</td>
<td>0.5 (16x)</td>
<td>0.125</td>
<td>0.008 (4x)</td>
<td>0.25</td>
<td>0.004</td>
<td>0.063</td>
<td>0.031</td>
<td>8.0</td>
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</tr>
<tr>
<td>A219/B</td>
<td>0.5 (16x)</td>
<td>0.125</td>
<td>0.008 (4x)</td>
<td>0.125</td>
<td>0.004</td>
<td>0.063</td>
<td>0.016</td>
<td>4.0</td>
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</table>

* NFLX = norfloxacin; CPFX = ciprofloxacin; ABPC = ampicillin; CTRX = ceftriaxone; CFTM = ceftopiram; IPM = imipenem; AZM = azithromycin; SPCM = spectinomycin; WT = wild type.
† The numbers in parentheses indicate the fold changes in MIC for a variant and its parent.

Table 2  Mutations in the quinolone resisting region of the gyrA and parC genes in N. gonorrhoeae. Nucleotides 265 to 291 of the gyrA gene coding for amino acids 89 to 97, and nucleotides 256 to 282 of the parC gene coding for amino acids 86 to 94 are shown. Dashes indicate sequence homology.
Discussion

Bellard et al.\(^{10}\) have recently generated a series of ciprofloxacin resistant mutants by exposing a N. gonorrhoeae isolate to increasing concentrations of ciprofloxacin. They have identified mutations in the gyrA and parC genes in these ciprofloxacin resistant strains. However, no mutation in the parC gene, without the simultaneous presence of a mutation in the gyrA gene has been identified. Their results have demonstrated that the serine-91 to phenylalanine substitution in the GyrA protein may be an important mutation in the development of gonococcal ciprofloxacin resistance. However, they have not examined whether the ciprofloxacin resistant variants displayed reduced accumulation of the drug in the cells.

In this investigation, we were able to generate several norfloxacin resistant mutants. All three wild type strains finally developed a mutation in the GyrA protein, although the mutation patterns differed. The MICs of norfloxacin for the variants A67/A, A219/A, and A219/B containing a single GyrA mutation were 16-fold higher than that for their parental isolates. The A69/A variant showing a markedly reduced norfloxacin accumulation in the cells, without mutations in the GyrA or ParC proteins, was also less sensitive to norfloxacin, with a 16-fold increase in the MIC, compared with the parent strain. The MIC of norfloxacin for the A69/B variant which contained a single GyrA mutation with reduced norfloxacin accumulation in the cells was 128-fold higher than for the parent strain. The MICs of norfloxacin for these five variants ranged from 0.063 to 0.5 \(\mu\)g/ml. These findings indicate that for the development of gonococcal fluoroquinolone resistance a mutation in the gyrA gene should be present and that a mutation leading to reduced drug accumulation also contributes to the development of fluoroquinolone resistance. Of the three parental isolates, only the A67/WT strain developed all the three known fluoroquinolone resistance mechanisms. The A67/A variant contained the Asp95→Tyr substitution in the GyrA protein and demonstrated 16-fold reduction in the MIC of norfloxacin compared with A67/WT. The A67/B variant acquired a Glu91→Val mutation in the ParC protein in addition to the pre-existing Asp95→Tyr mutation in GyrA, and was remarkably resistant to norfloxacin as evidence by a norfloxacin MIC of 4.0 \(\mu\)g/ml which was 256 times that of the A67/WT strain. In addition, this isolate showed significantly decreased accumulation of norfloxacin. These data suggest that a mutation in the parC gene with the simultaneous presence of a mutation in the gyrA gene contributes a high level of fluoroquinolone resistance in gonococci with decreases in accumulation in cells having an additional but lesser effect.

In Gram negative bacteria such as \(E\) coli, fluoroquinolone resistant mutants with a reduction in OMP F (porin) have been isolated.\(^{21,22}\) In the present study, to determine whether alterations occurred in the structural elements of the outer membrane of the fluoroquinolone resistant \(N\) gonorrhoeae variants, we compared the OMP profiles of the variants with their parental isolates. No differences were identified in OMP profiles between the fluoroquinolone resistant variants with or without decreased norfloxacin accumulation and the parental isolates. Therefore, it is suggested that the gonococcal variants have other mechanism such as active efflux system across the inner membrane\(^{27}\) leading to reduced norfloxacin accumulation in the cells.

15. Hooper DC, Wolfson JS, Roza MA, Ng EY. Genetics and regulation of outer membrane protein by quinolone resistance loci \(nfb\), \(nfc\), and \(xeb\). Antimicrob Agents Chemother 1992;36:1151–4